

International Journal of Frontline Research in Chemistry and Pharmacy

Journal homepage: https://frontlinejournals.com/ijfrcp/ ISSN: 2945-4824 (Online)





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# Successive infusion of Mncl<sub>2</sub> in order to analyze the image produced by MRI machines

Habibu Ahmad Ibrahim<sup>1,\*</sup>, Yusuf Abubakar Maitama Hotoro<sup>1</sup>, Abubakar Sani Garba<sup>2</sup> and Bushra Khan<sup>3</sup>

<sup>1</sup> Department of Physics, Kano University of Science and Technology Kano, Nigeria.

<sup>2</sup> Department of Physics, College of Art and Remedial Studies Kano, Nigeria.

<sup>3</sup> Department of Radiology, Imaging Sharda University and Sharda Hospital Utter Pradesh, India.

International Journal of Frontline Research in Chemistry and Pharmacy, 2022, 01(01), 032–036

Publication history: Received on 27 February 2022; revised on 02 April 2022; accepted on 04 April 2022

Article DOI: https://doi.org/10.56355/ijfrcp.2022.1.1.0005

### Abstract

The use of MRI machines is a fast growing in the field of Medical Imaging. MRI machine operates on the principal of Nuclear Magnetic Resonance Imaging which can be explained by Quantum Mechanics or Classical Mechanics. Nuclear Magnetic Resonance Imaging is a process by which atomic nuclei containing protons and neutrons which are all inform of a magnetic field absorb and re-emit electromagnetic radiation. The energy of the emitted electromagnetic radiation is usually at a certain resonance frequency, which depends upon the strength of the magnetic field and the magnetic properties of the atoms. The resonance frequency produced is similar to radio frequency RF radiation, which when observed it produce a fine spectrum of anatomical structure of objects. MRI machines are currently used in hospitals and clinics to take the anatomical structures of the human body. And a wide variety of artifacts is routinely encountered on the images produced by this Machines. Manganese is a metal that was proposed to be use in NMRI machines, but due to it's toxic nature and it is a heavy metal, it is not use at all. But we are able to produce metallic chloride of Mn<sup>2+</sup> which over comes the toxicity of metal and behaves like a metallic Salt in the human body. However, we are able to successfully infuse MnCl2 into a Rat and a Rabbit due to its unique properties in order to analyse the images produced by a MRI Machine. Conclusively, we are able to observe that MnCl<sub>2</sub> improves the images quality produced by the MRI Machine.

Keywords: Nuclear; Resonance; Toxicity; MRI

### 1. Introduction

In the year 1973, Paul Lauterbur present a seminar paper that was titled "Image formation by induced local interactions: examples employing nuclear magnetic resonance" which was later published. In the paper, the basic concepts of MRI was extensively explained. This concepts was used later in modern MRI. However, in the content of his paper a new type of medical imaging was proposed which depends one on the magnetic resonance frequency of water hydrogen atoms when an externally magnetic field gradient of same frequency is applied. At the time of Lauterbur's work, the notion to image water distribution seemed to be limited for medical applications because water density varied by only a small degree in tissues. However, by that time there were already results that demonstrated that magnetic resonance relaxation times of water were different in different tissues and might be altered by pathology [2]. Lauterbur used the well know ability of the paramagnetic ion, Mn<sup>2+</sup> to alter the longitudinal relaxation time of water [1]. Today there are over 80 million MRI exams per year and approximately 25% rely on adding contrast [3].

However, there have been much interest to use  $Mn^{2+}$  to serve as a contrast agent in taking the anatomical images of animals. Whenever this method is developed it can be use or MRI of human beings. Though, the ability of  $Mn^{2+}$  to serve as an MRI contrast agent gives more possibilities for using Manganese Chloride (MnCl<sub>2</sub>) to improve the images produced. A lot of research works has been performed showing the use of  $Mn^{2+}$  in order to improve the quality of images

\* Corresponding author: Habibu Ahmad Ibrahim

Department Of Physics, Kano University of Science And Technology Kano, Nigeria.

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produced by MRI machines. However, we have reviewed this recent work using  $MnCl_2$  and we are able to analyse the quality of images produced by MRI Machines.

## 2. Material and methods

#### 2.1 Materials

The materials used in this research work was a 3 Tesla MRI Machine that was constructed by Siemens. This machine has an active shielded super-conducting magnet with an operational field strength of 3 Tesla ( $\pm 10\%$ ) suitable for high-resolution structural imaging, functional imaging, diffusion imaging and spectroscopy. It also consist of a gradient system, with active shielding in X, Y, Z and other planes, that is capable of simultaneously achieving a maximum gradient strength of 80 milli Tesla/meter with a slew rate of 200 Tesla/meter/sec along each axis with a 100% duty cycle for full FOV. The MRI machine consists of a hardware and software that is capable to perform proton spectroscopy and multinuclear spectroscopy along with optimized sequences and software for post processing and evaluations. We also prepare MnCl<sub>2</sub> in a liquid form at room temperature and pressure. A suitable digital balance was also used. Few calibrated syringes where also involved in the research work. Finally, we are able to use a rat and rabbit of 350 grams and a rabbit of 1.35 kilograms.

#### 2.2 Methodology

The method we used was a successive infusion of the MnCl<sub>2</sub> with a syringe and then injecting it into the two samples; which were the rat and the rabbit. However, we first all prepare the MnCl<sub>2</sub> in its liquid form at room temperature. We then test the MnCl<sub>2</sub> to make sure that it has completely changed to a metallic salt, by mixing it with a small fragment of concrete H<sub>2</sub>SO<sub>4</sub> acid. Which after mixing the metallic salt and acid they eventually produced a base. This result confirmed that the MnCl<sub>2</sub> can be as metallic salt just like NaCl<sub>2</sub> (common salt). After we have prepared the MnCl<sub>2</sub>, we then use a calibrated syringe. By putting 5ml of the MnCl<sub>2</sub> into the syringe. We then take our first sample which the rat and we measured it's weight using a digital balance. After the measurement, we then infused or rather inject the MnCl<sub>2</sub> into the sample and we recorded the time of the infusion. After 10 minutes of the infusion, we then introduced the sample into the MRI Machine and take the anatomical structure of rat brain. This method was also repeated to rabbit only that 10ml of MnCl<sub>2</sub> was administered into the rabbit because of its weight.

After 24 hours of the successive infusion of MnCl<sub>2</sub> into the rat and the rabbit. They were also administered into the MRI Machine to take their anatomical structure of the brain respectively. However, this method was successfully continued for 3 days. We are able to analyse the images produced by MRI Machine.

### 3. Results and discussion

Generally the use of MnCl<sub>2</sub> as a MRI contrast agent depend on giving or infusion of MnCl<sub>2</sub> and also monitoring its distribution in the tissues of our samples. But our major concern was to monitor how the MnCl<sub>2</sub> affected our samples before infusion and after infusion. Conclusively, after infusion MnCl<sub>2</sub>, our samples were not affected at all before and after the infusion. However, at each instance of the infusion we took MRI images of the brain of our samples. Below is the MRI image of a rat brain after successive infusion of 5ml of MnCl<sub>2</sub> at an interval 4 hours respectively.



Figure 1 The MRI images of the rat brain after successive infusion of MnCl2.

However, after 24 hours the MRI image of the rat brain was taken and it was compared with the previous images. But the image produced was far better in contrast compared with the later images. Conclusively, after comparing the MRI images, it reveals that the successive infusion of MnCl<sub>2</sub> improves the image quality produced.



Figure 2 The MRI image of a rat brain after 24hours of successive infusion of MnCl2.

Similar procedure was conducted for the rabbit and the MRI images shows that the image quality was improved. This can be seen below.



Figure 3 The MRI images of the rabbit brain after successive infusion of MnCl2



#### Figure 4 The MRI image of a rabbit brain after 24hours of successive infusion of MnCl2

#### 4. Conclusion

Generally, most of MRI images when they are produced are often encountered with different artefacts. A lot of research have been done and more research are still on trying to find out methods on how to minimise this artefacts. In our research work we have use MnCl<sub>2</sub> which has served as a common salt and we have administered it dose into a rat and a rabbit by successively infusing it into the rat and the rabbit respectively. The images that were produced after successive infusion of MnCl<sub>2</sub> have a better contrast compared to those MRI images produced before the infusion of MnCl<sub>2</sub>. However, we have also gradually monitored how the rat and the rabbit behaves before and after the successive infusion of MnCl<sub>2</sub>.

Conclusively, the rat and the rabbit were not affected at all after infusing the dose of MnCl<sub>2</sub> into their bodies. Finally, after the successive infusion of MnCl<sub>2</sub> into our samples and their MRI images were taken, analysed and compared. The images produced have a nicer contrast. This means that the successive infusion of MnCl<sub>2</sub> improves the quality of images produced by MRI Machines.

#### References

- [1] Lauterbur PC. Image formation by induced local interactions: examples employing nuclear magnetic resonance. nature. 1973 Mar;242(5394):190-1.
- [2] Damadian R. Tumor detection by nuclear magnetic resonance. Science. 1971 Mar 19;171(3976):1151-3.
- [3] de Haën C. Conception of the first magnetic resonance imaging contrast agents: a brief history. Topics in Magnetic Resonance Imaging. 2001 Aug 1;12(4):221-30.
- [4] Li Y, Huang T, Carlson E, Melov S, Ursell P, Olson J, Noble L, Yoshimura M, Berger C, Chan P. Nat. Genet. 11, 376– 381 Wedler, F. and Denman, R. (1984) Curr. Top. Cell Regul. 1995; 24: 153.
- [5] Lee and Koretsky . Biochemistry. 1979; 18: 3642-3646.
- [6] Morita H, Ogino T, Seo Y, Fujiki N, Tanaka K, Takamata A, Nakamura S, Murakami M. Neurosci. Lett. 2002; 326: 101-104.
- [7] Hu T, Pautler R, MacGowan G, Koretsky A. Magn Resonan. Med. 2001; 46: 884-890.
- [8] Pautler R, Silva A, Koretsky A. Magn. Resonan. Med. 1988; 40: 740-748.
- [9] Watanabe T, Michaelis T, Frahm J. J Magn Resonan. Med. 2001; 46: 424-429.
- [10] Van Der Linden A, Verhoye M, Van Meir V, Tindemans I, Eens M, Absil P, Balthazart J. J. Neuroscience. 2002; 112: 467-474.
- [11] Saleem K, Pauls J, Augath M, Trinath T, Prause B, Hasjikawa T, Logothetis N. Neuron. 2002; 34: 685-700.
- [12] Lin Y. MRI of the rat and mouse brain after systemic administration of MnCl 2. Thesis, Carnegie Mellon University, Pittsburgh, PA. 1997.
- [13] Watanabe T, Natt O, Boretius S, Frahm J, Michaelis T. Magn. Resonan. Med. 2002; 48: 852-859.
- [14] Aoki I, Lin-Wu Y, Silva A, Lynch R, Koretsky A. Neuroimage. 2004; 22: 1046-1059.
- [15] Lauterbur P, Mendonca Dias M, Rudin A. Augmentation of tissue water proton spin-lattice relaxation rates by in vivo addition of paramagnetic ions. In: Frontiers of Biological Energetics, Dutton, P., Leigh, J. and Scarpa, A. eds. Academic Press, NY. 1978; 1: 752-759.
- [16] Hollis D, Bulkley B, Nunnally R. Clin. Res. 1978; 26: 240A.
- [17] Baum L, Wolf GL. Cardiovascular toxicity and tissue proton T1 response to manganese injection in the dog and rabbit. American Journal of roentgenology: Volume 141. 1983:193.
- [18] Kang YS, Gore JC. Studies of tissue NMR relaxation enhancement by manganese. Dose and time dependences. Investigative radiology. 1984 Sep 1;19(5):399-407.

- [19] Saeed M, Higgins C, Geschwind J, Wendland M. Eur. Radiol. 2000; 10: 310-318.
- [20] Koretsky A, Detre J, Williams DS, Ho C. Unpublished observation. 1988.
- [21] Nunnally RL, Hollis DP. Adenosine triphosphate compartmentation in living hearts: a phosphorus nuclear magnetic resonance saturation transfer study. Biochemistry. 1979 Aug 1;18(16):3642-6.
- [22] VANDER ELST LU, Colet JM, Muller RN. Spectroscopic and metabolic effects of MnCl2 and MnDPDP on the isolated and perfused rat heart. Investigative radiology. 1997 Oct 1;32(10):581-8.
- [23] Castro CD, Koretsky AP, Domach MM. NMR-Observed phosphate trafficking and polyphosphate dynamics in wildtype and vph1-1 mutant Saccharomyces cerevisae in response to stresses. Biotechnol Prog. 1999;15(1):65-73.
- [24] Federle M, Chezmar J, Rubin D, Weinreb J, Freeny P, Semelka R, Brown J, Borello J, Lee J, et al. J. Magn Resonan. Imag. 2000; 12: 186-197.
- [25] Bremerich J, Saeed M, Arheden H, Higgins CB, Wendland MF. Normal and infarcted myocardium: differentiation with cellular uptake of manganese at MR imaging in a rat model. Radiology. 2000 Aug;216(2):524-30.